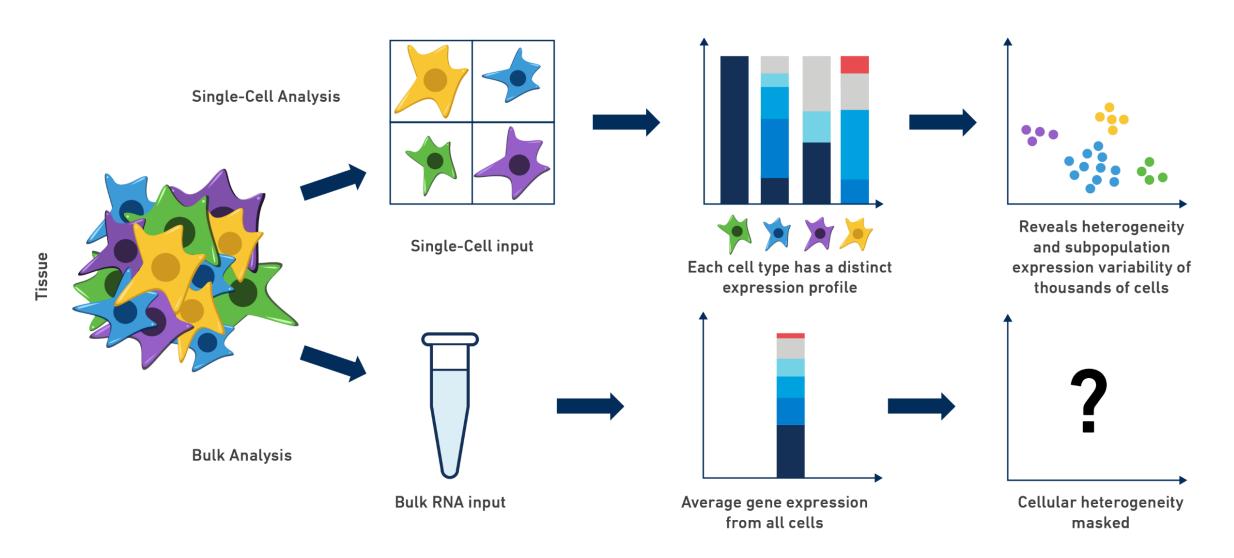
Single Cell Transcriptomics – a simple walkthrough

Adwait Sathe, PhD

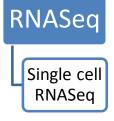
McDermott Center Bioinformatics Lab <u>adwait.sathe@utsouthwestern.edu</u>



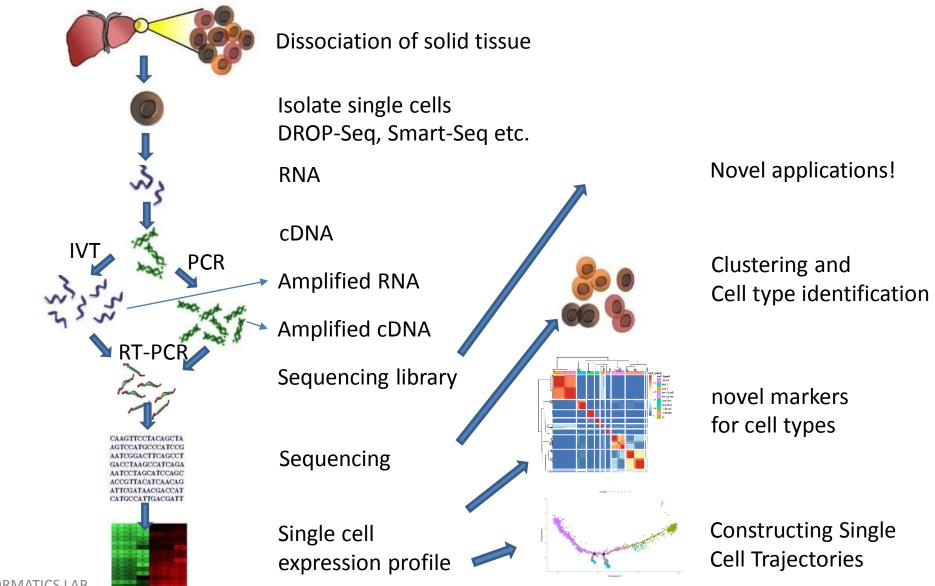




https://www.10xgenomics.com/solutions/single-cell/



Single cell RNA Sequencing Workflow



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RNASeq

Single cell RNASeq

ScRNAseq methods

Single cell RNA Sequencing methods comparison

Chromium, 10X

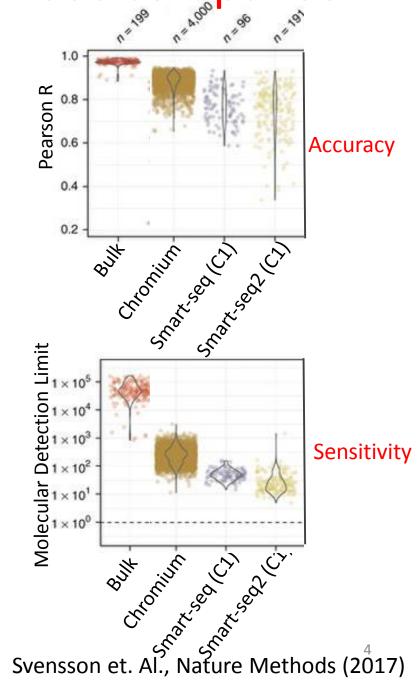
- UMI unique molecular identifiers
- 3' counting (only 3' of transcript sequenced)
- 8-10 bp UMI
- 10X barcoded gel beads
- Throughput (number of cells) 10^4 – 10^5

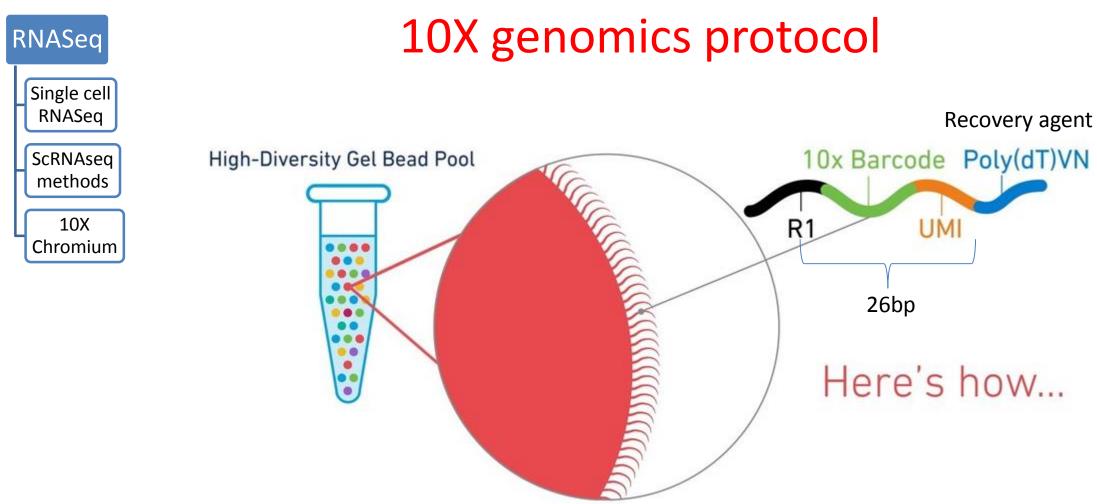
Smart-seq/C1 microfluidic platform

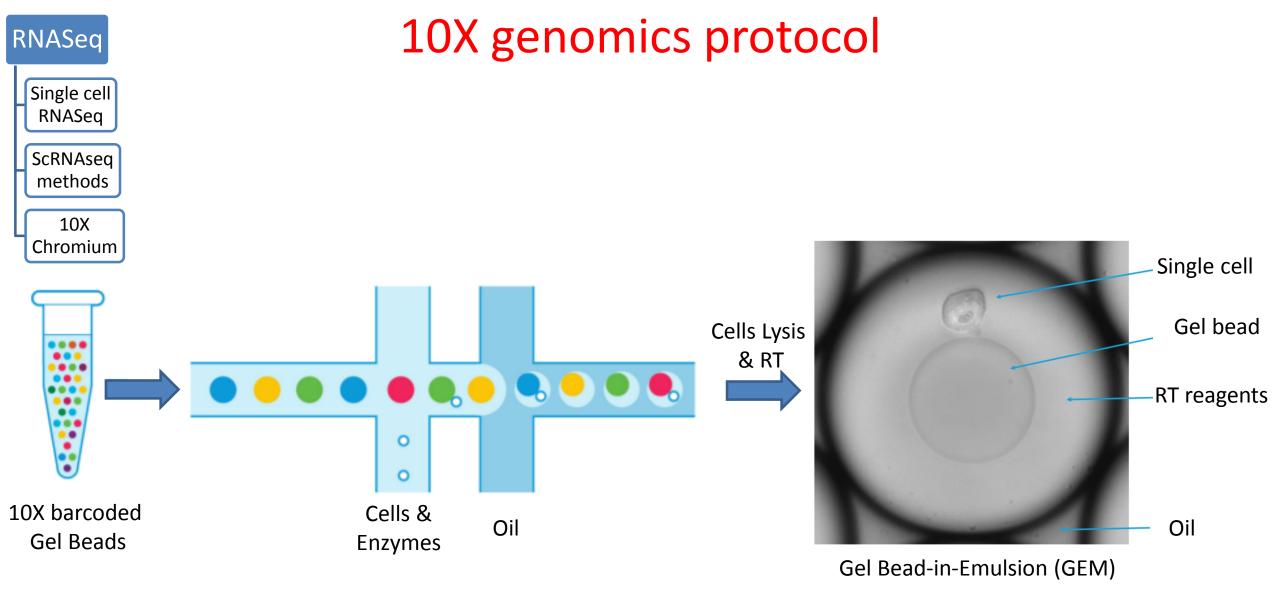
- Full length
- No UMI
- Fluidigm C1
- Throughput (number of cells) 10^2 – 10^3

Smart-seq2 most genes per cell among methods

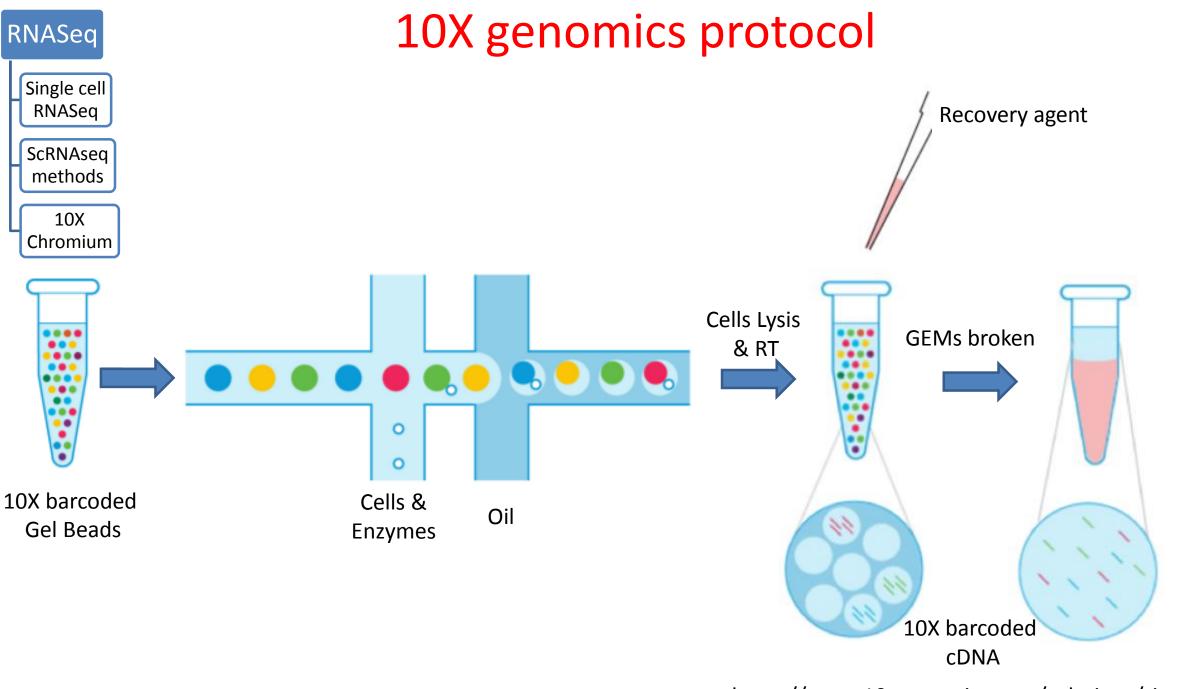
- Full-length
- No UMI
- FACS
- Throughput (number of cells) 10^2 – 10^3





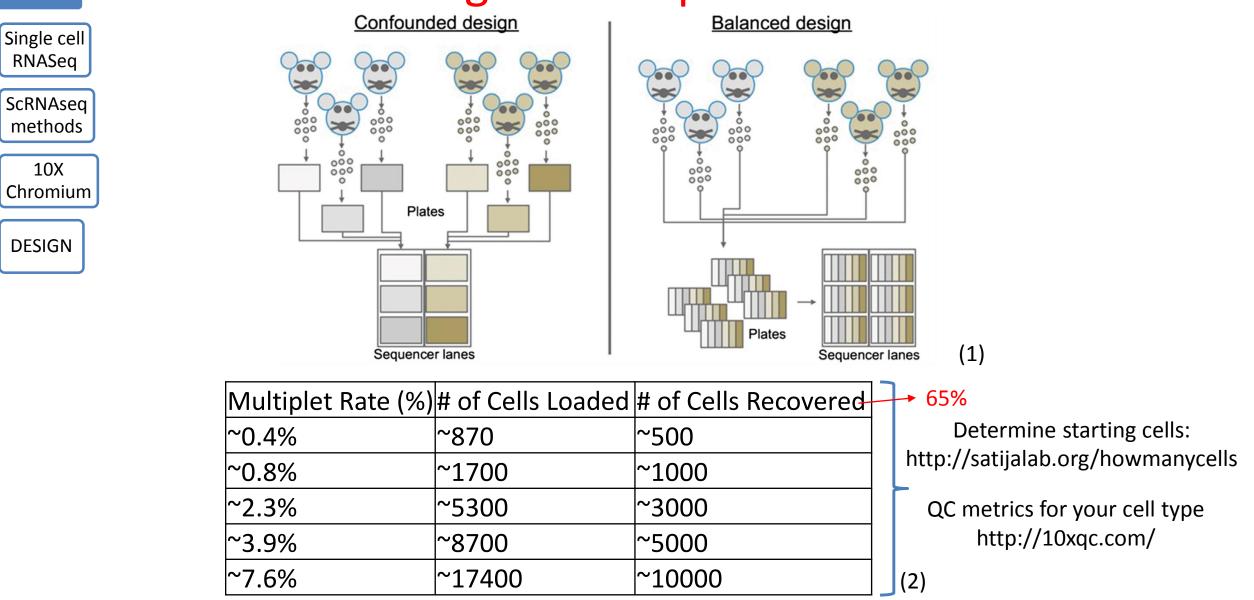


https://www.10xgenomics.com/solutions/single-cell/



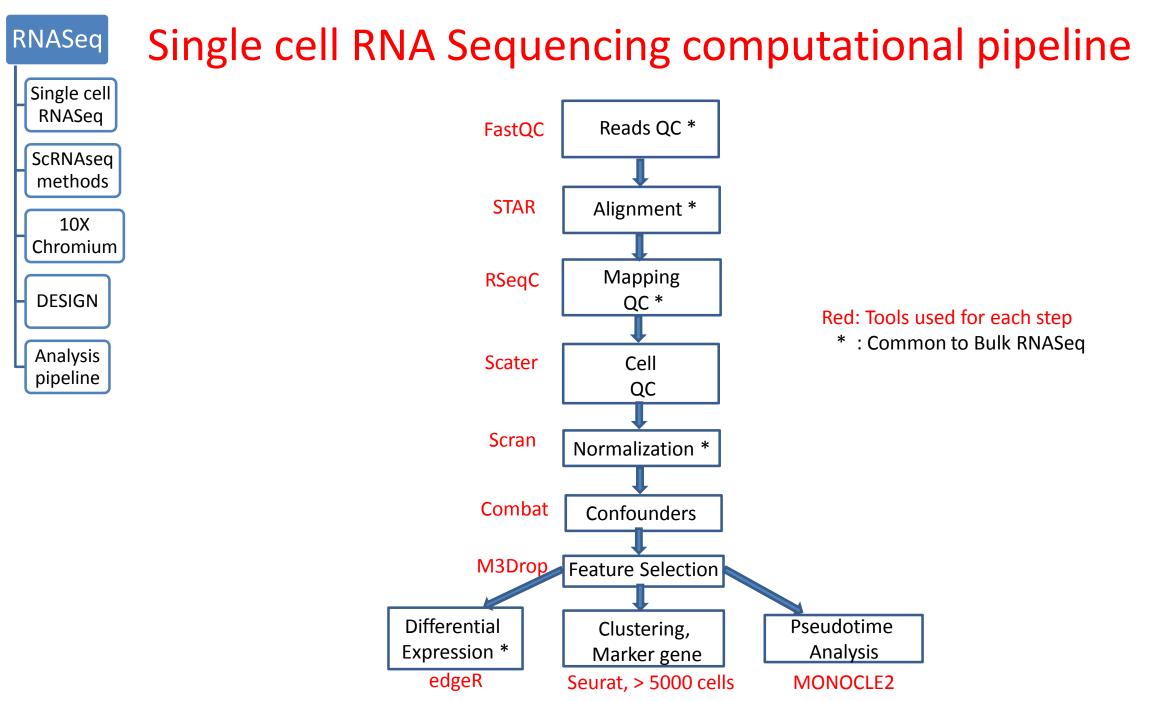
https://www.10xgenomics.com/solutions/single-cell/

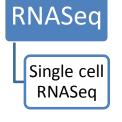




(1) Jeanette Baran-Gale et. al., Brief Funct Genomics, 2017. doi:10.1093/bfgp/elx035
 (2) https://www.10xgenomics.com/solutions/single-cell

RNASeq

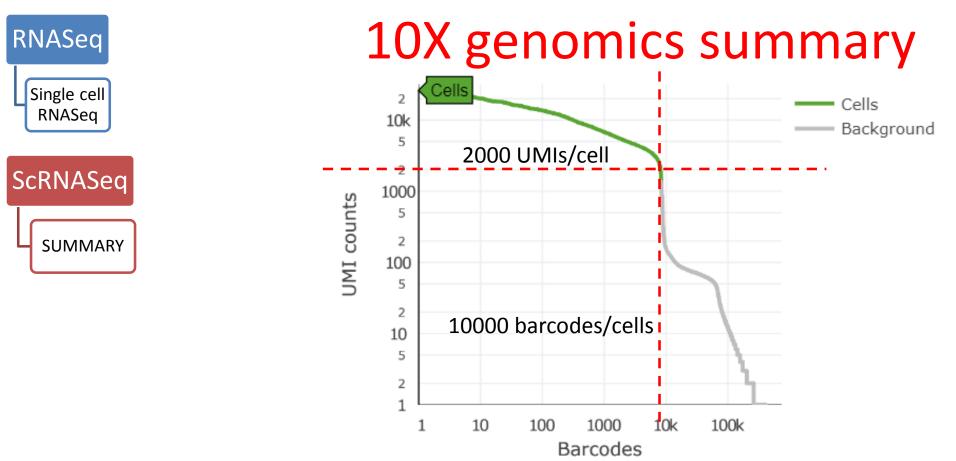




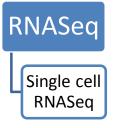
ScRNASeq

Let us carry out a single cell RNASeq experiment

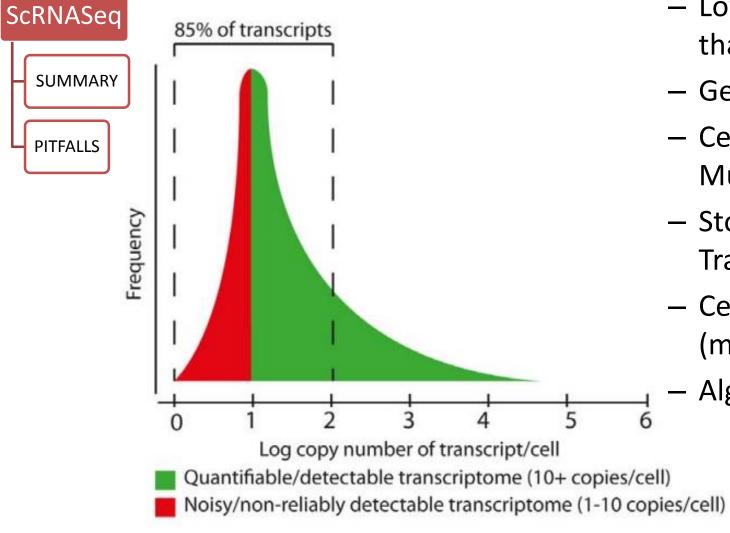
- Peripheral blood mononuclear cells (PBMCs) from a healthy donor. PBMCs are primary cells with relatively small amounts of RNA (~1pg RNA/cell).
- Sequenced on Illumina Hiseq4000
- 26bp read1 (16bp Chromium barcode and 10bp UMI), 98bp read2 (transcript), and 8bp I7 sample barcode



Parameter	PBMC Observed	PBMC Recommended
Estimated Number of Cells/Barcodes	8381	500-10,000
Fraction Reads in Cell	93.10%	
Mean Reads per Cell	93,552	50,000
Median Genes per Cell	1297	1000
Total Genes Detected	21,425	
Median UMI counts per cell	4084	



Potential problems

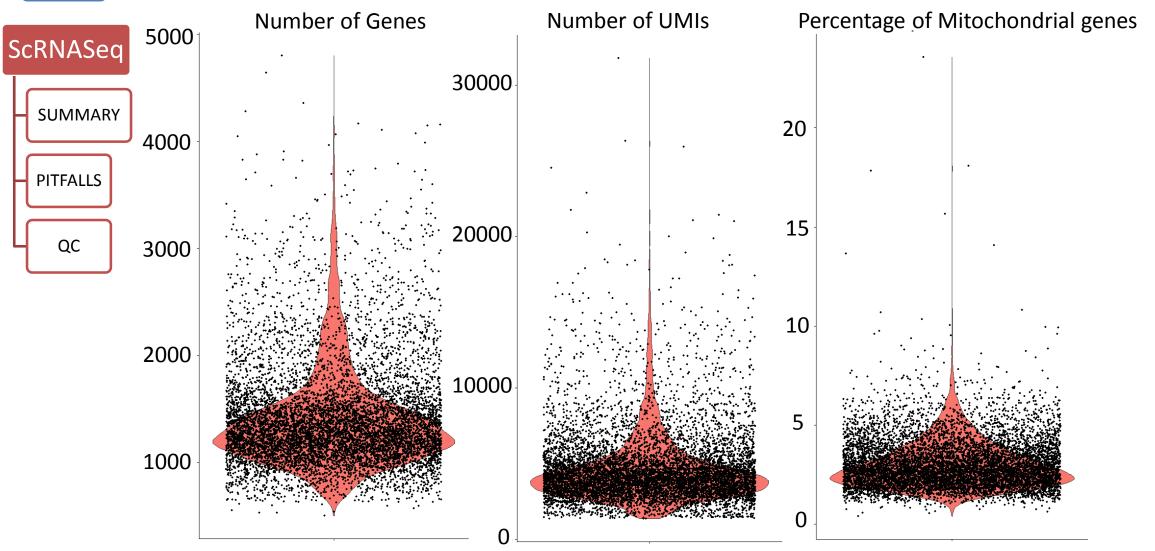


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- Low amplification efficiency, typically less than 10%
- Gene dropout rates
- Cell quality: Live/dead, Missing cells, Multiple cells
- Stochastic: cell cycle phases, Transcriptional bursting
- Cell Capture rate ≠ population frequency (multiples/gel beads, empty beads)
- Algorithm development

Macaulay and Voet, PLOS Genetics, 2014. Computational Methods for Analysis of Single Cell RNA-Seq Data, Ion Măndoiu, University of Connecticut.

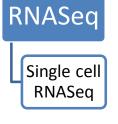
Exploring the Quality Control required

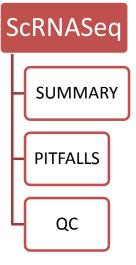


PBMC sample

RNASeq

Single cell RNASeq

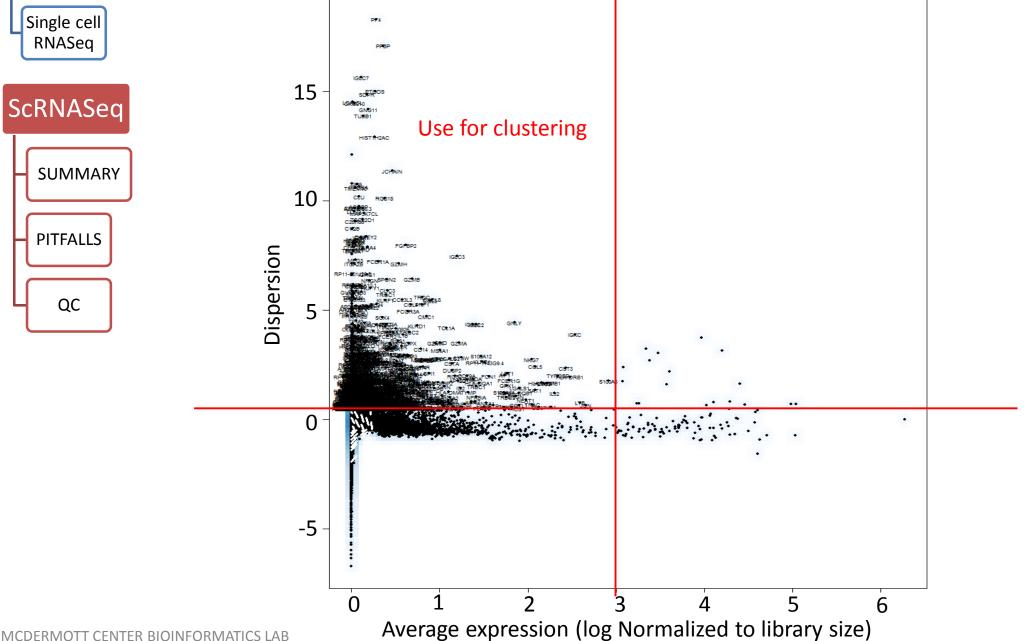




Clustering

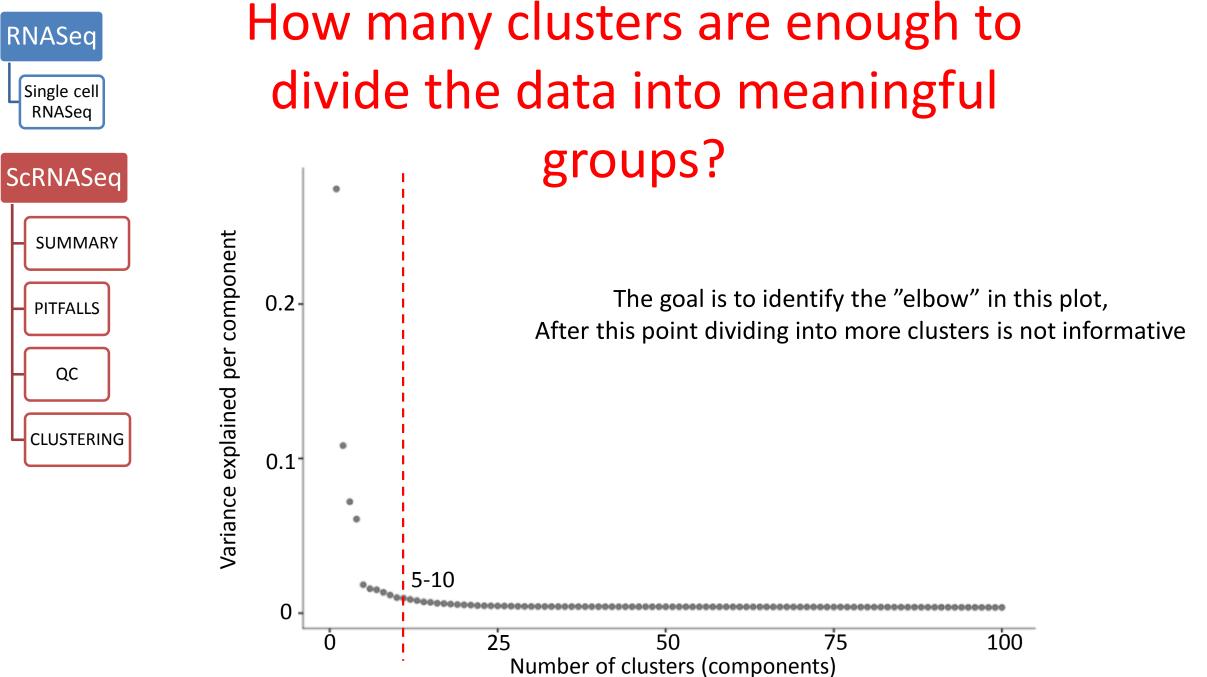
- Determine a subset of genes to use for clustering; this is because not all genes are informative, such as those that are lowly expressed.
- The approach is to select gene based on their average expression and variability across cells
- We scale the data and remove unwanted sources of variation (technical, cell cycle stage, batches etc.)

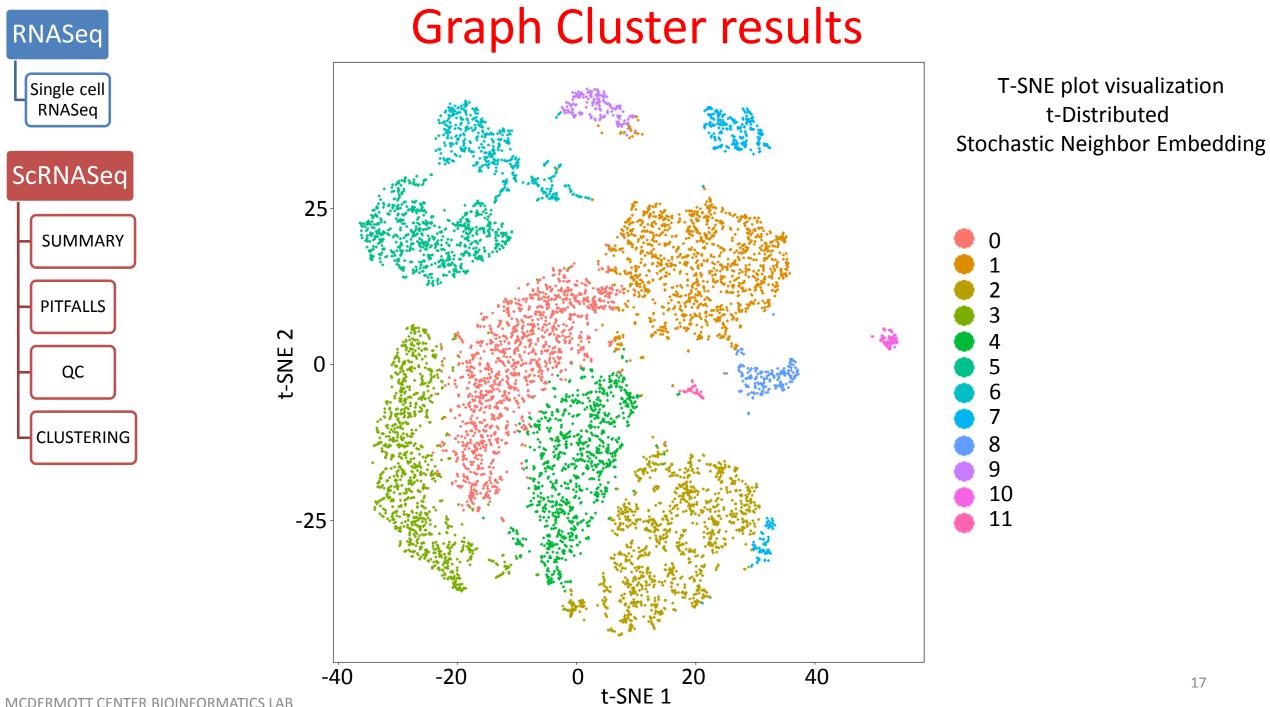
Mean vs Variation: Detection of variable genes RNASeq

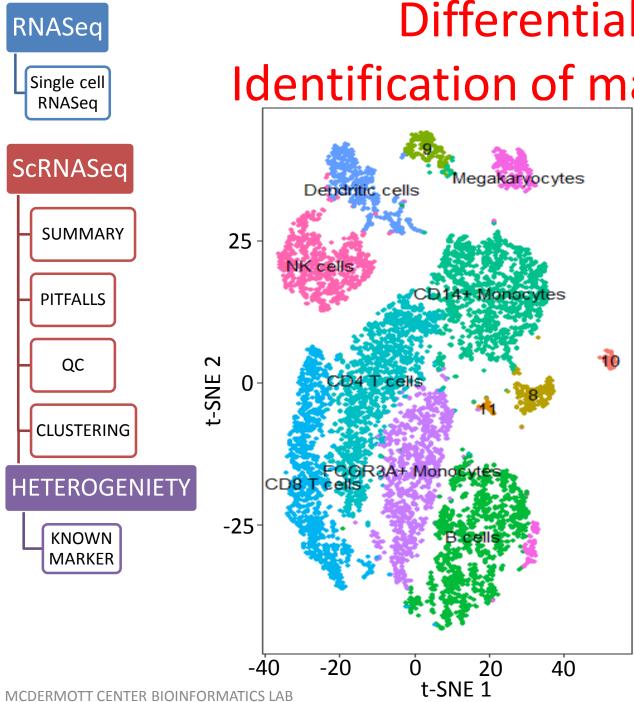


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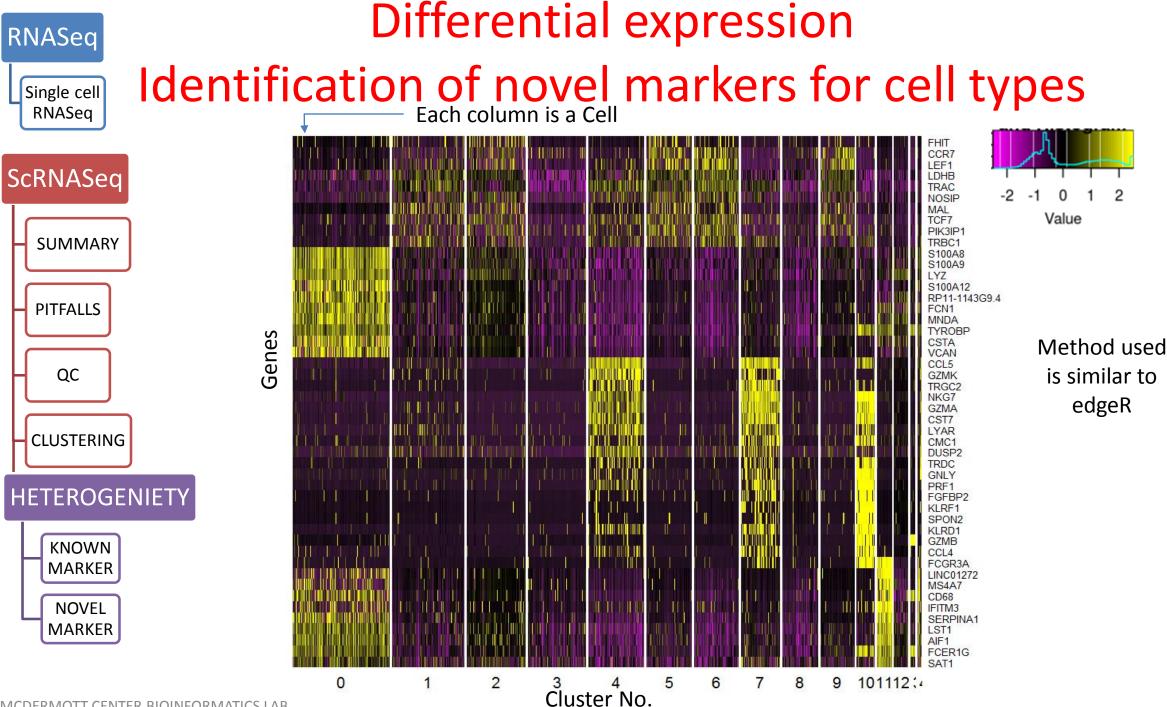


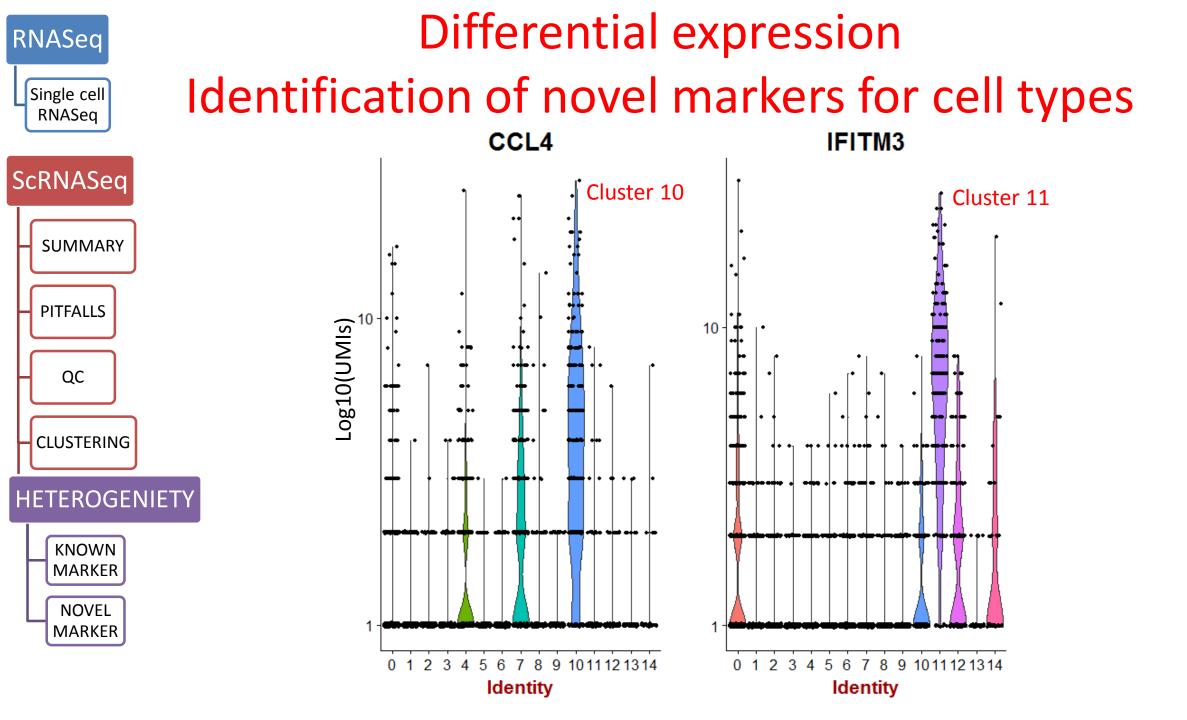


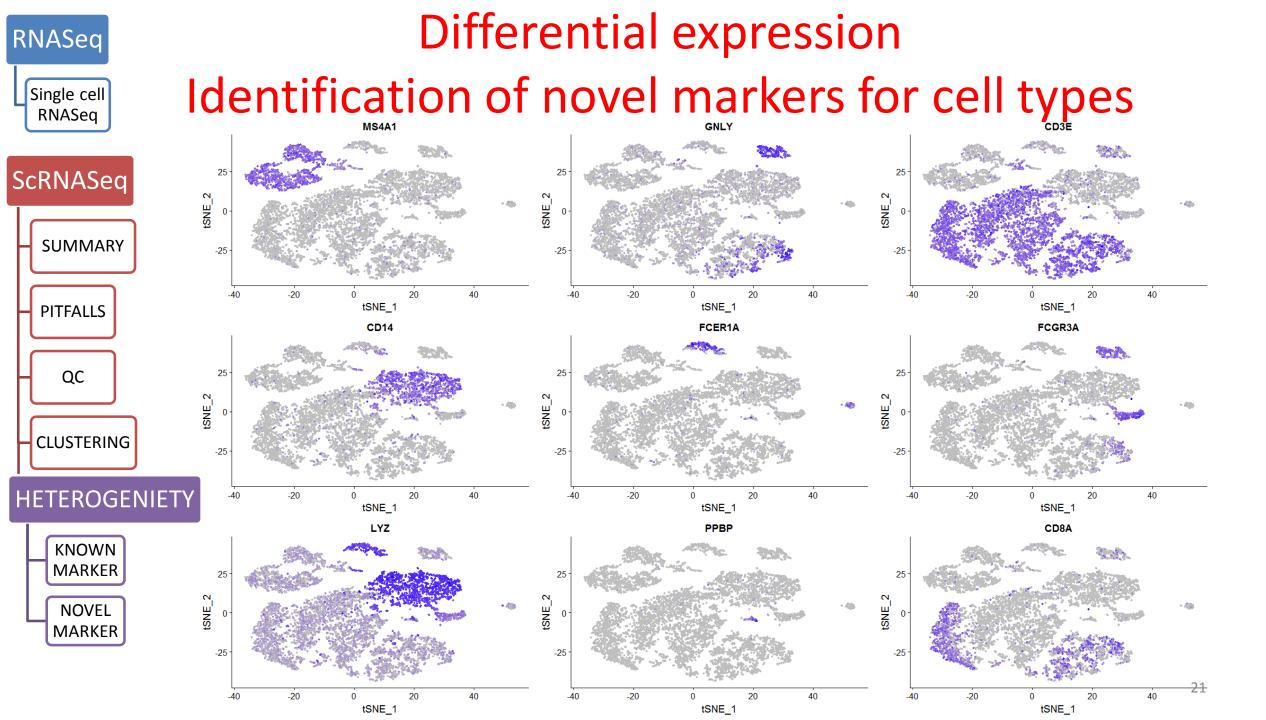
Differential expression Identification of markers for cell types

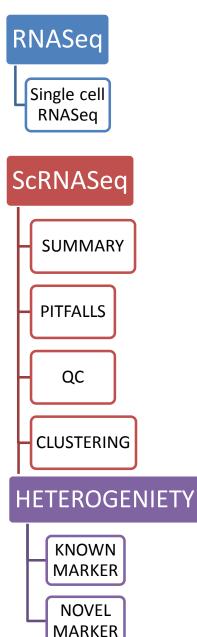
Supervised – PBMC known markers

Markers	Cell Type	
IL7R	CD4 T cells	
CD14, LYZ	CD14+ Monocytes	
MS4A1	B cells	
CD8A	CD8 T cells	
FCGR3A, MS4A7	FCGR3A+ Monocytes	
GNLY, NKG7	NK cells	
FCER1A, CST3	Dendritic Cells	
РРВР	Megakaryocytes	









Cloupe browser further exploration

Acknowledgements

McDermott Center Bioinformatics Lab McDermott Center NGS core team

Chao Xing, PhD - Director Mohammad Kanchwala Ashwani Kumar

Workflow used is based on:

 Seurat: Macosko, Basu, Satija et al., Cell, 2015 (Updated approach: Combining dimensional reduction with graph-based clustering)
 Monocle: Xiaojie Qiu, Andrew Hill, Cole Trapnell et al (2017)
 Computational Methods for Analysis of Single Cell RNA-Seq Data, Ion Măndoiu, University of Connecticut

THANK YOU